

Development of a Clinical-Grade Meganuclease for Allogeneic CAR T Cell Production

Janel Lape, Victor Bartsevich, Caitlin Turner, Mara Davis, Keith Wetzel, Jochen Genschel, Audrey Brown, Melissa Samo, Milloni Chhabra, Aaron Martin, Jeff Smith



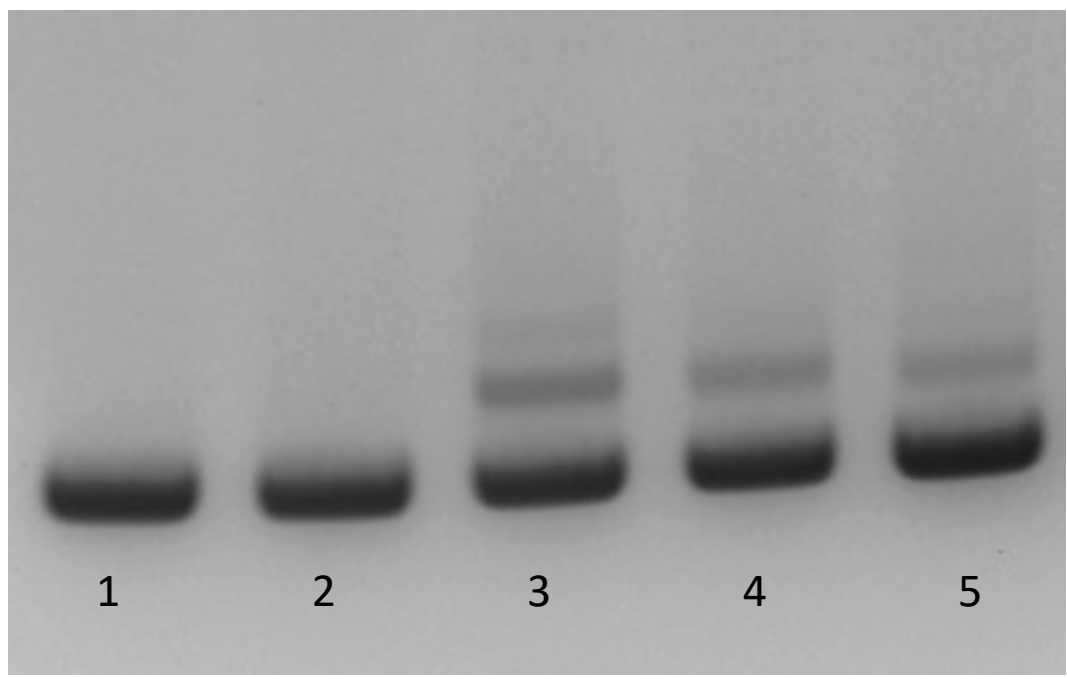
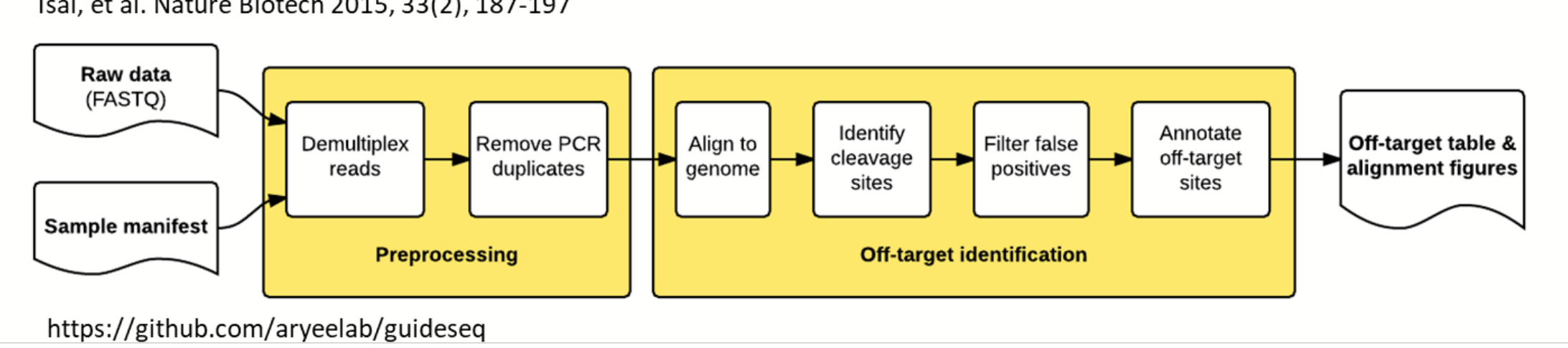
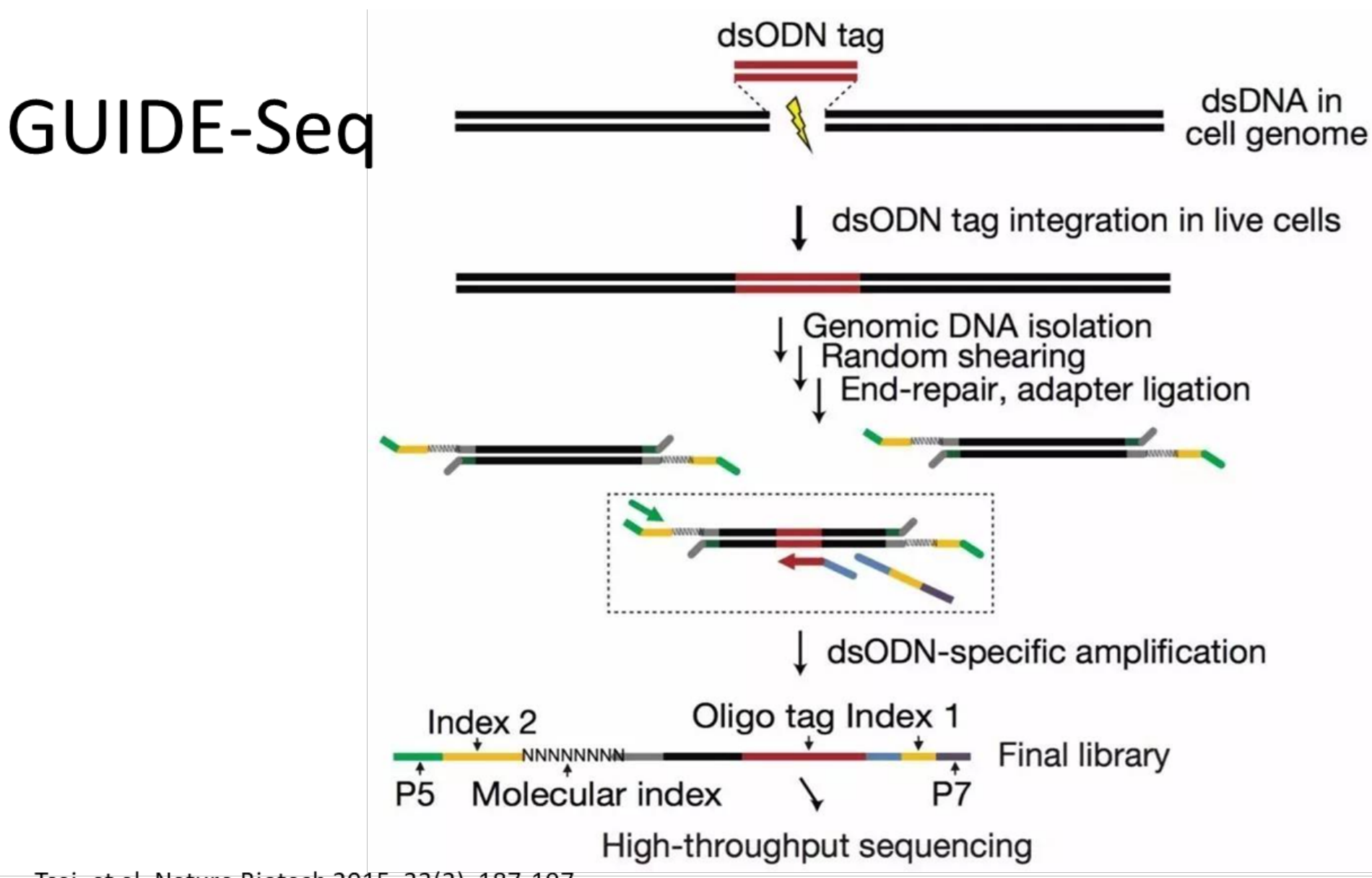
ABSTRACT

Chimeric Antigen Receptor T cells (CAR Ts) are revolutionizing the treatment of hematologic malignancies. Autologous (patient-derived) therapies, though effective, are challenging to produce. Allogeneic (donor-derived) CAR T cells may be an attractive alternative if they can be modified to prevent graft-vs-host disease (GvHD). We previously reported an efficient genome editing strategy to produce allogeneic CAR T cells by targeting the insertion of a CAR transgene directly into the native TRAC locus using an engineered meganuclease and an AAV donor template (MacLeod, et. al., 2017). The resulting cells are CAR+ and do not elicit GvHD by virtue of having the native T cell receptor gene knocked-out by the CAR transgene.

To better understand the extent and impact of nuclease activity on the CAR T cell product, we developed a high-sensitivity assay for off-target cutting that takes advantage of the 3' overhangs generated by meganucleases. Based on this analysis, we were able to engineer a second-generation nuclease with greatly reduced off-target activity. By comparing the performance of the optimized nuclease to the parent, we found a surprisingly robust relationship between nuclease specificity and CAR T cell performance. These results suggest that monitoring and control of off-target nuclease activity is critical to optimizing the overall fitness and function of gene edited CAR T cells.

Oligo capture parallels GUIDE-Seq in wet lab procedure with changes at a few key points. Instead of using a blunt-ended double stranded oligo for insertion, we use a double stranded oligo with randomized, protected, 4bp overhangs at the 3' ends. A new data processing pipeline has been customized for meganucleases, including changes in target site identification based on the single chain structure.

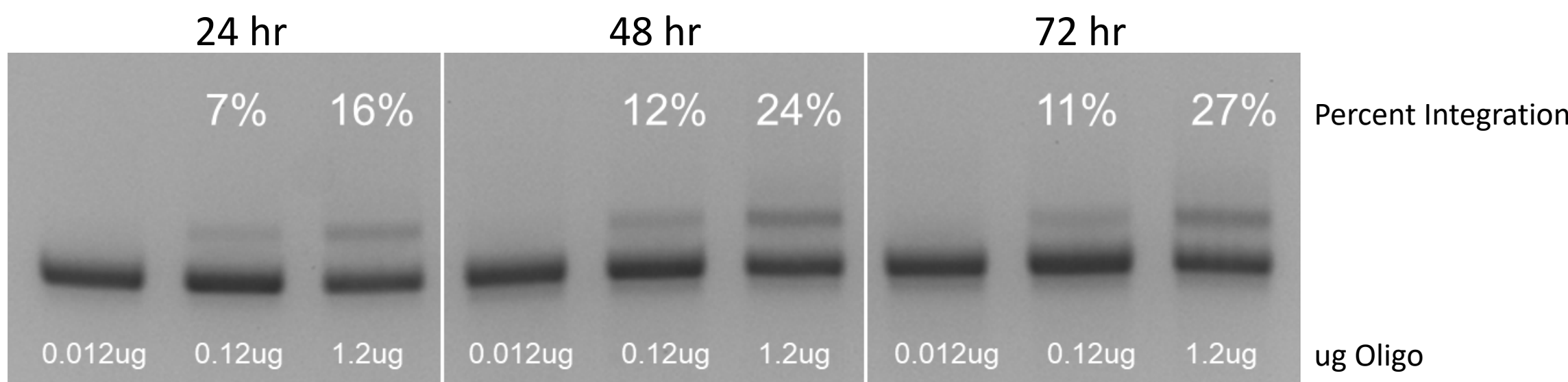
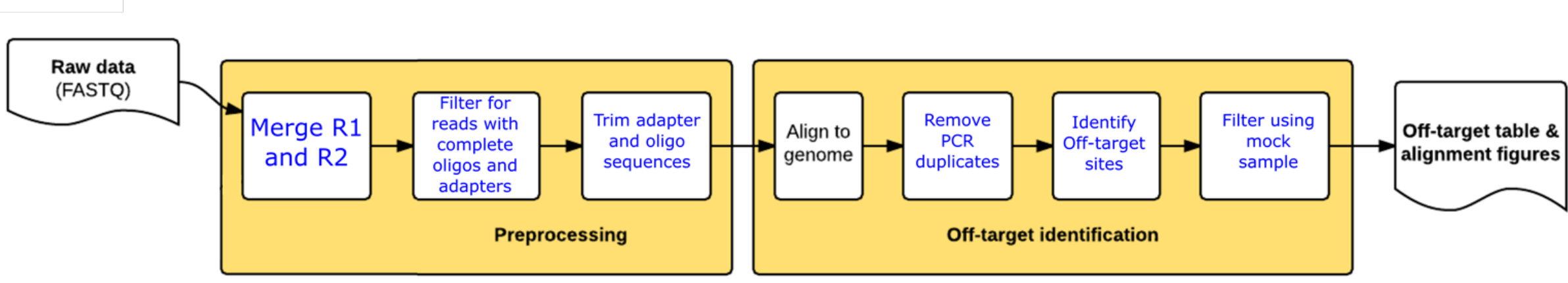
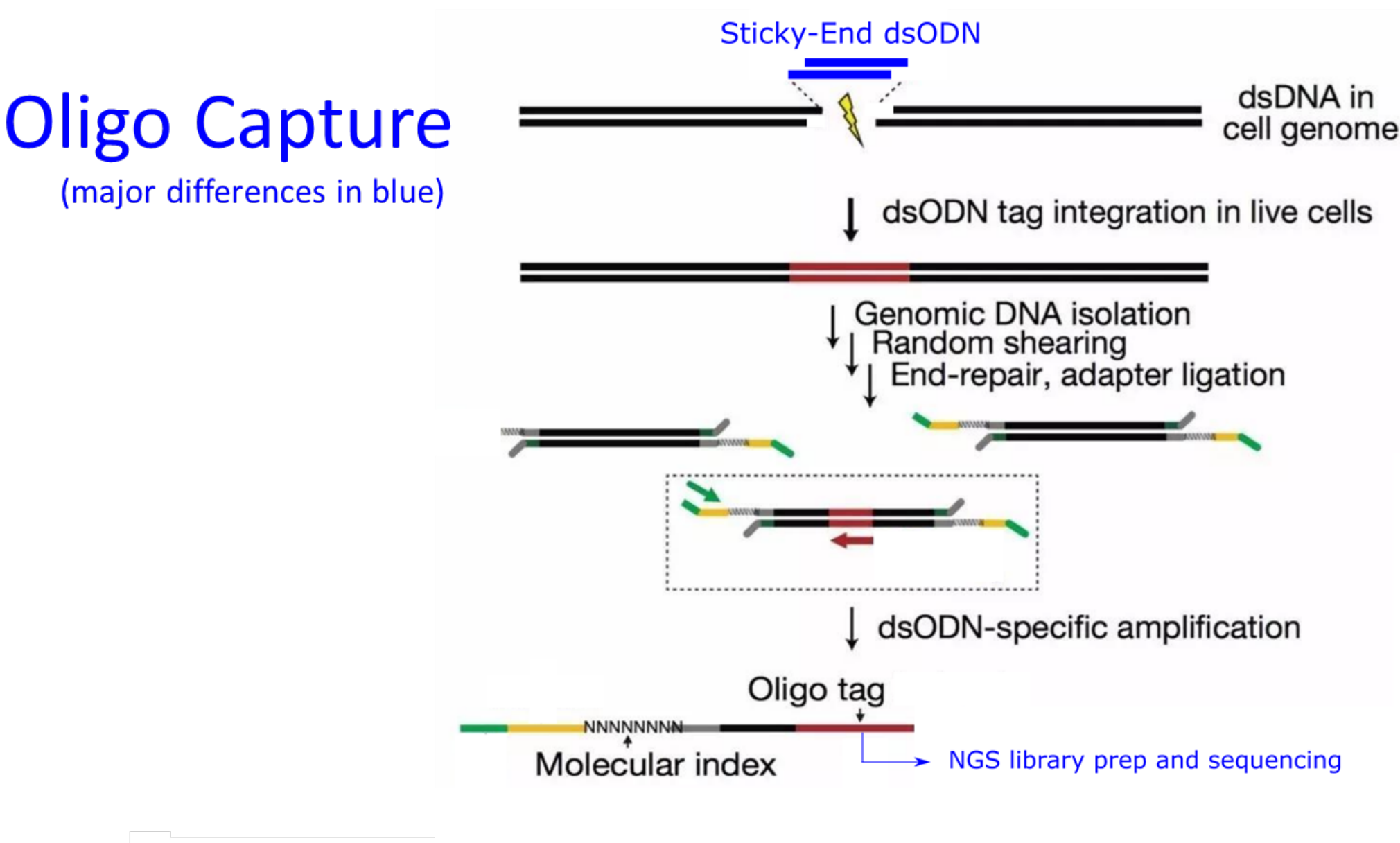
GUIDE-Seq



	Integration
1 Mock	0%
2 Nuclease only	0%
3 Nuclease + oligo with site specific overhangs	19%
4 Nuclease + oligo with random overhangs	14%
5 Nuclease + oligo with blunt ends	8%

Adding a randomized 4bp 3' overhang increases the integration of the oligo at the intended target site from 8% to 14%.

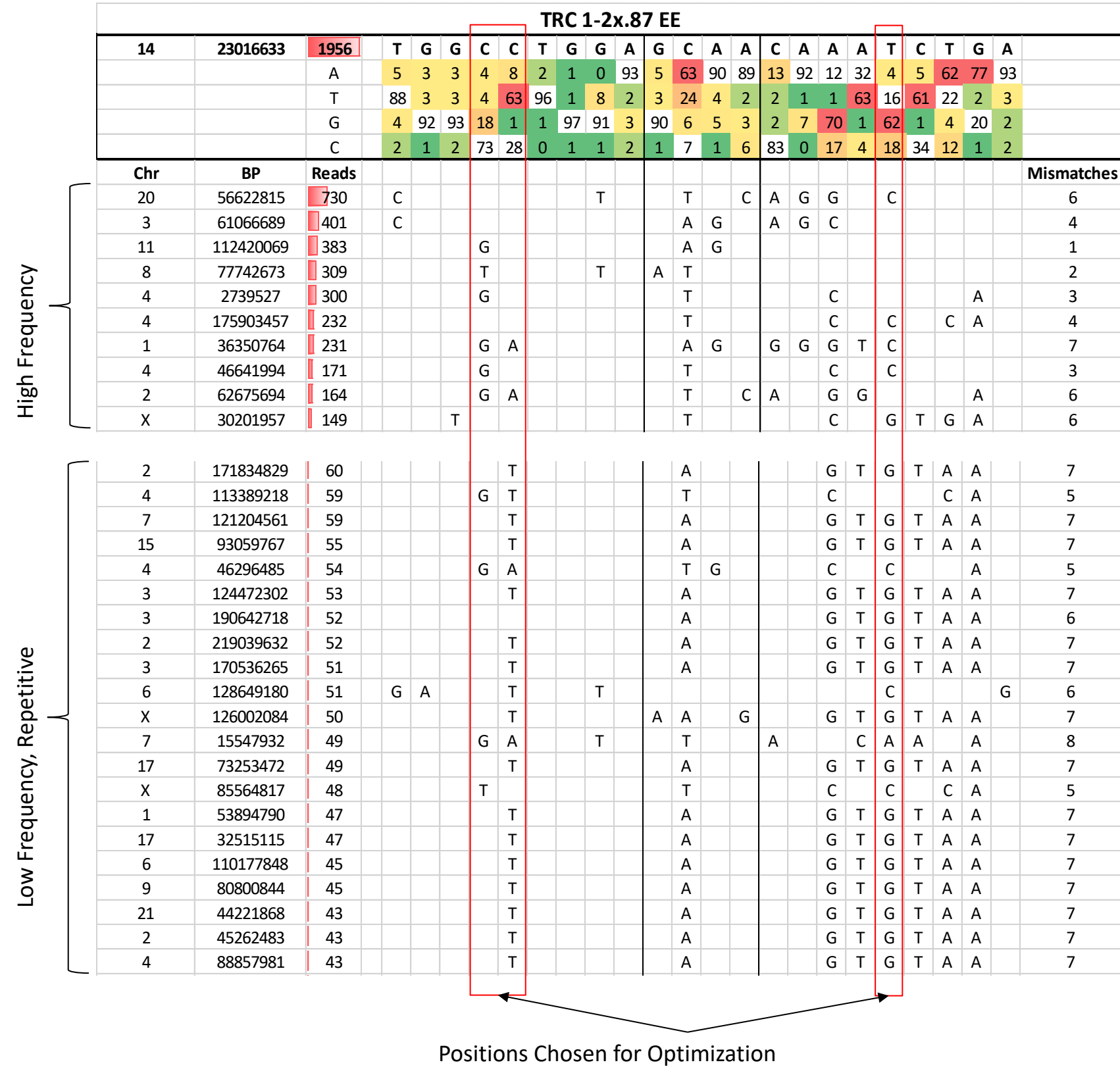
Oligo Capture



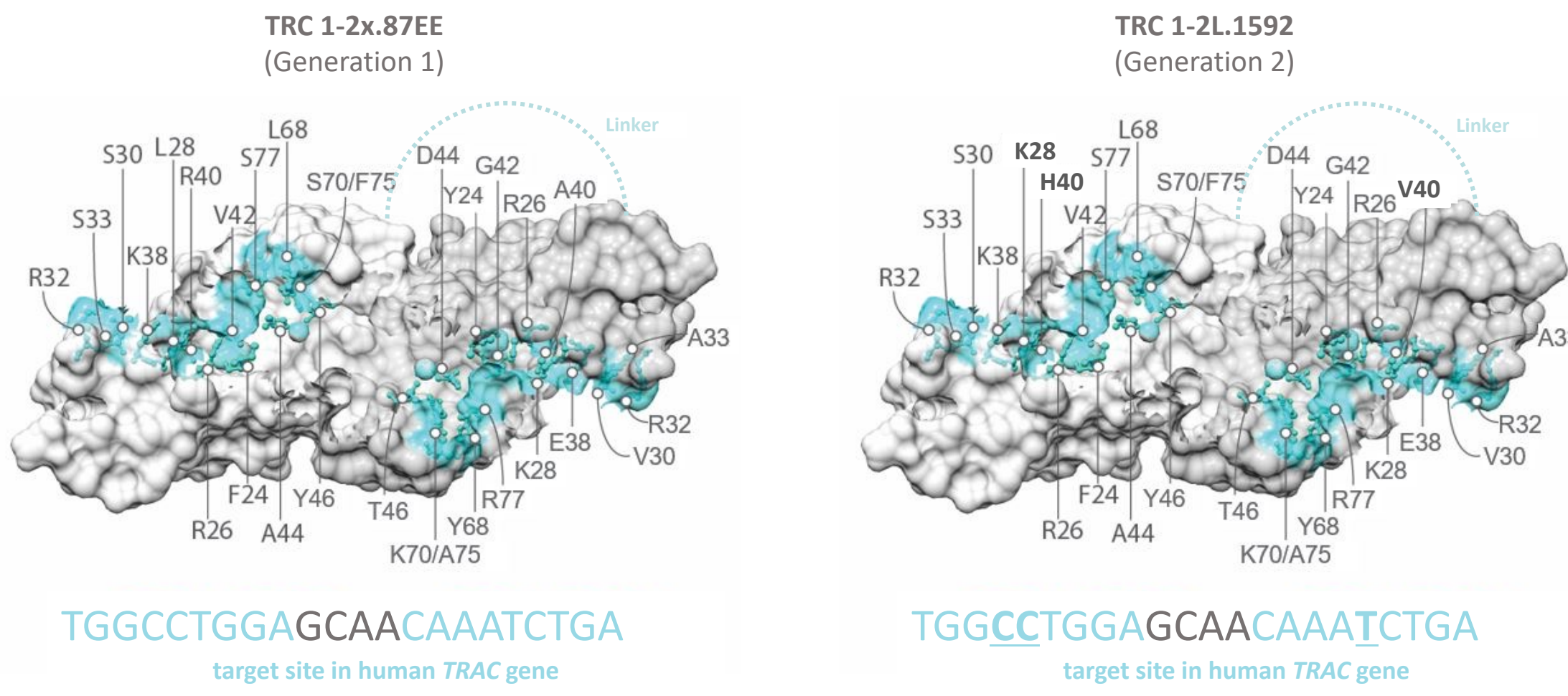
Integration scales with amount of oligo electroporated into the cells and peaks around 48 hours. No significant increase in integration occurs between 48 and 72 hours post-electroporation.

RESULTS

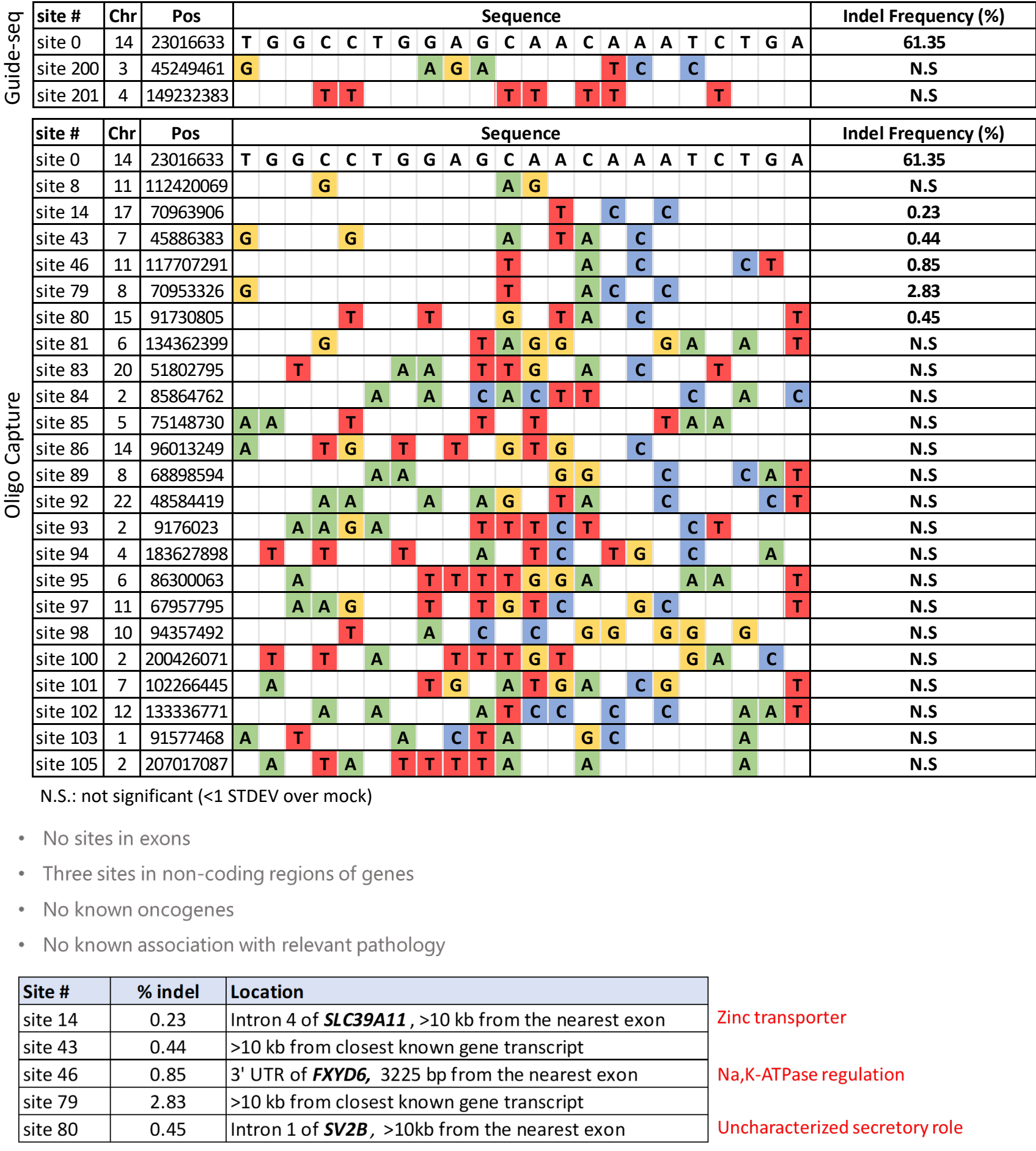
Oligo capture performed on the TRC 1-2x.87 EE nuclease identified two types of off-target sites. The first type were high frequency hits: sites that were cut many times resulting in a large number of reads. The second type were low frequency repeats: sites that were cut a few times and had lower numbers of reads but with a target site sequence that occurred multiple times in the genome.



Three positions in the target sequences were chosen for optimization and a second generation enzyme was produced (TRC 1-2L.1592). The second generation nuclease differs from the original at only 3 amino acid positions (**designated in bold**).

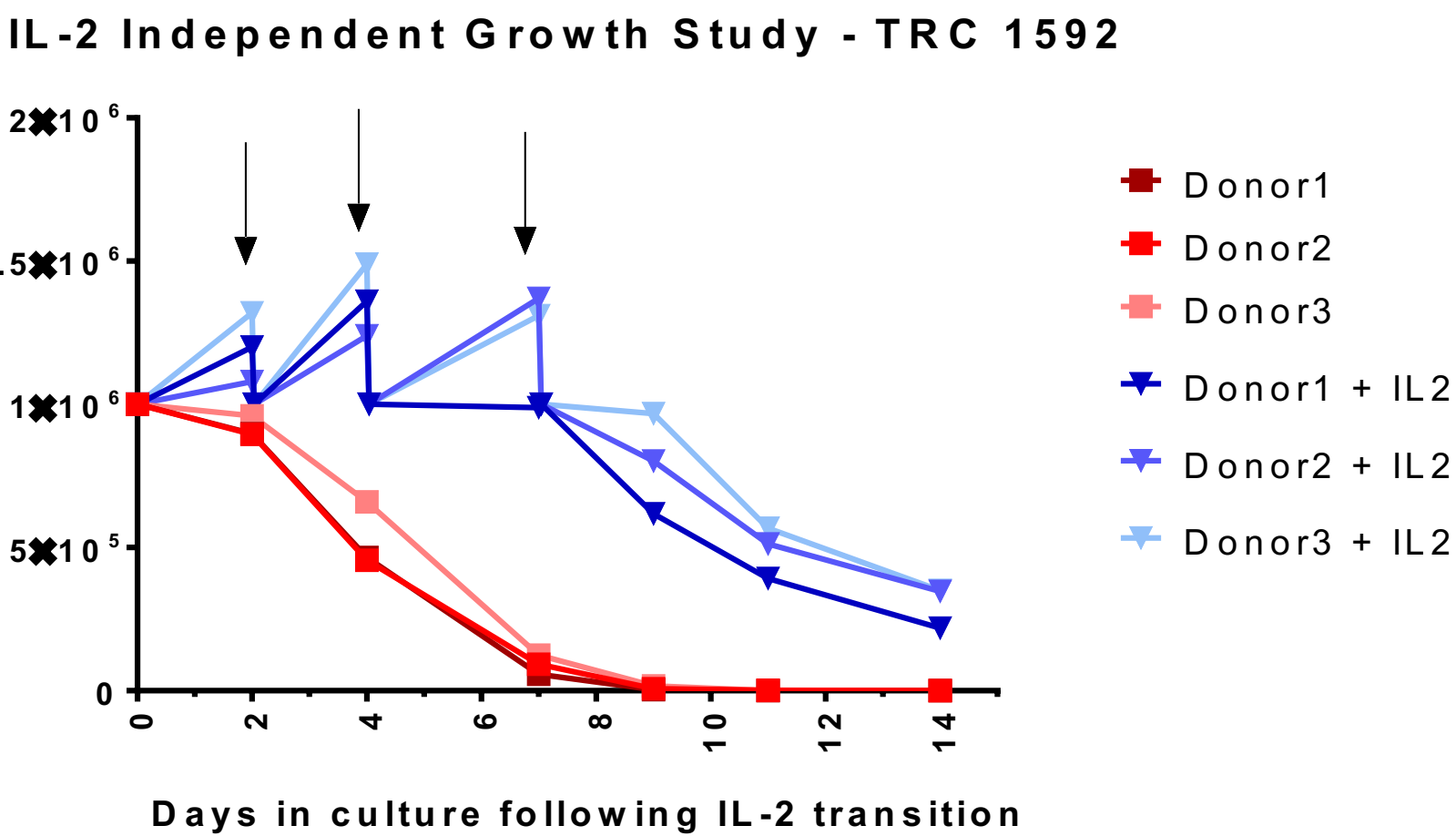
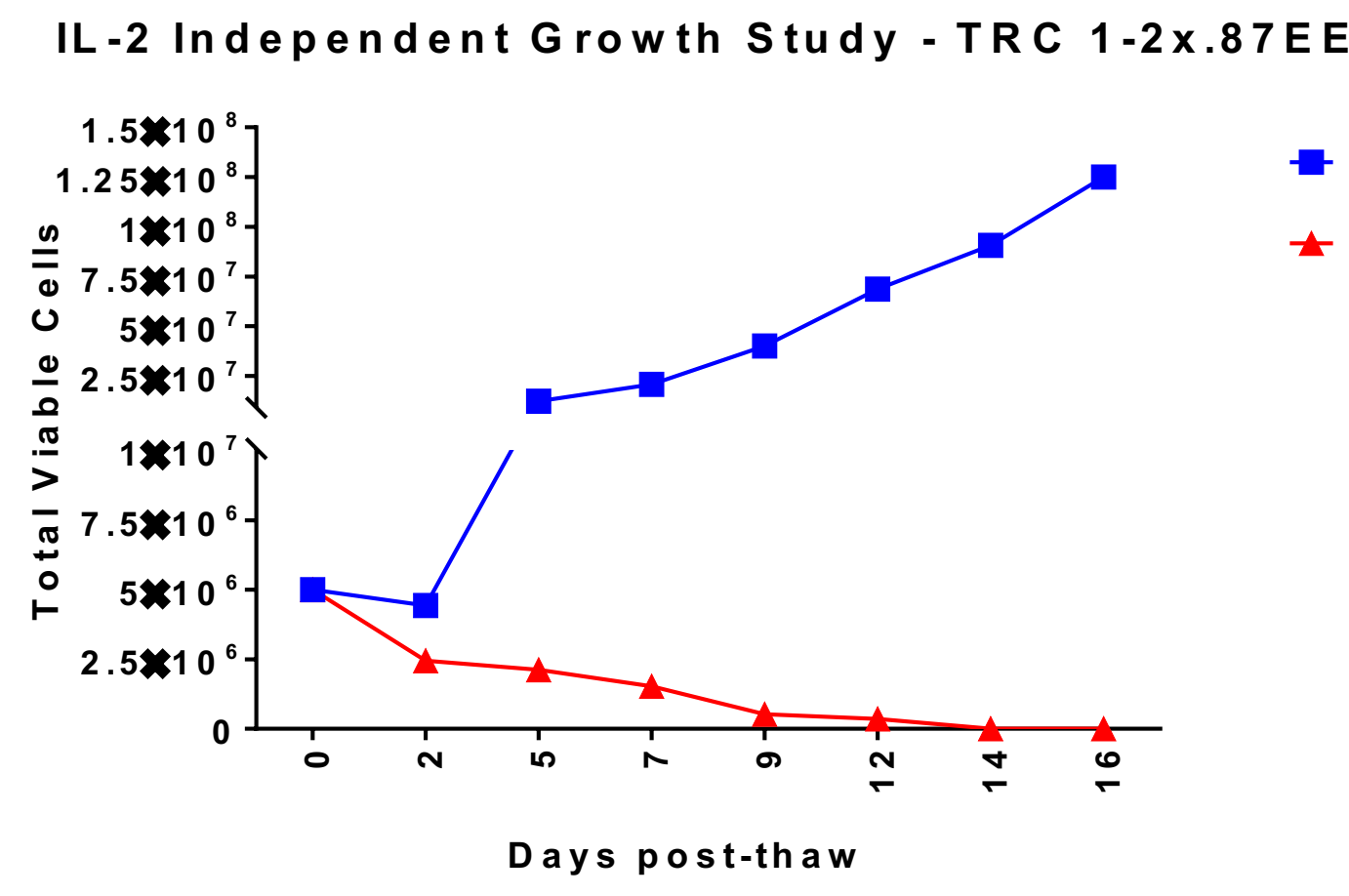
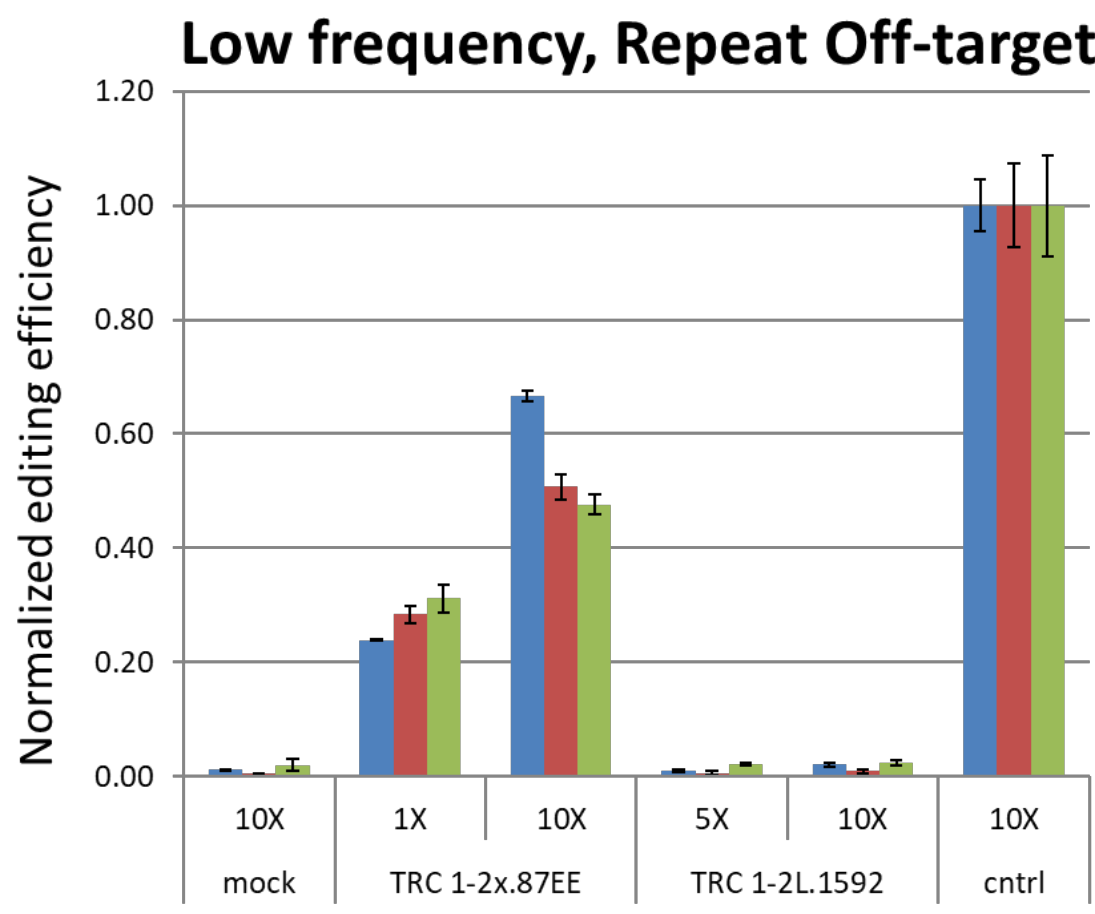
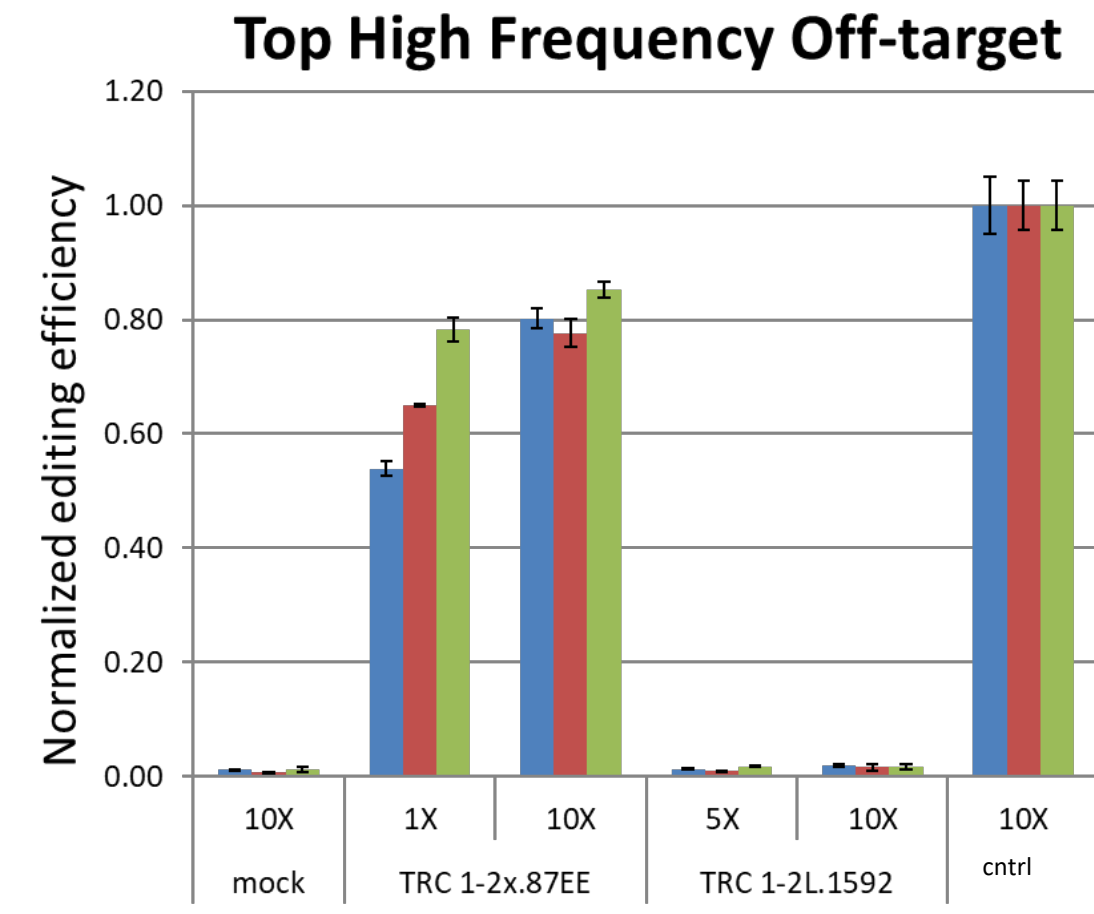


Analysis of TRC 1-2L.1592 by Guide-seq produced only two off-target sites, both of which were shown to be false positives by targeted indel analysis. Oligo capture identified a further 47 targets, 5 of which produced indels above the mock background.



Reporter assays showed that the second generation nuclease did not cut either the top high frequency off-target site or the low frequency, repetitive off-target site.

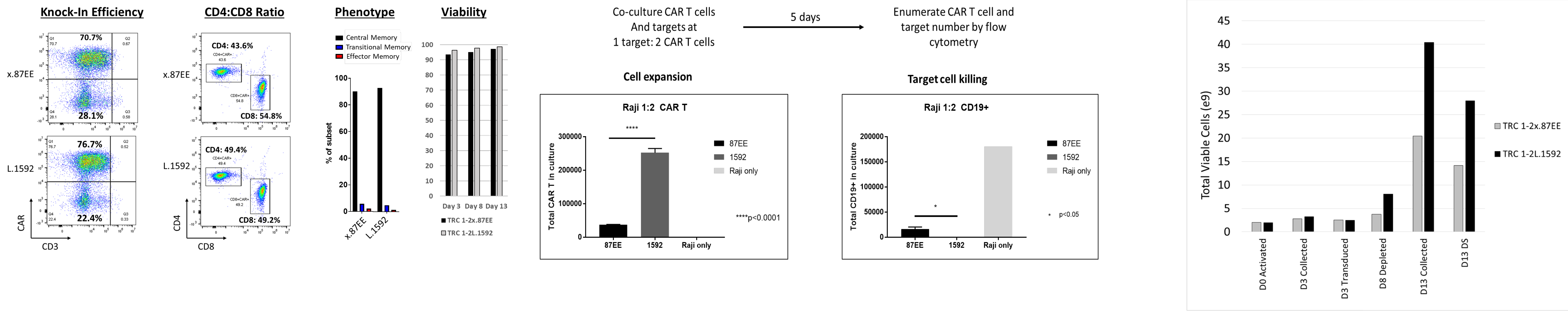
CAR T cells produced with either the first or second generation nuclease fail to survive in the absence of exogenous IL-2 indicating that no neoplastic transformation has occurred.



The optimized TRC 1-2L.1592 showed a better knock-in efficiency, CD4:CD8 ratio, and phenotype. Viability was unaffected.

CAR T cells produced using TRC 1-2L.1592 expanded 5x better than those made with TRC 1-2x.87EE and killed target cells significantly better.

Significant increases were seen in the number of total viable cells produced in a standard production run.



Conclusions

Guide-seq has become the gold standard for identification of off-target genome editing using CRISPR, but several changes are needed to allow discovery of possible off-targets generated by a meganuclease. Using our customized method, we were able to identify and eliminate multiple off-target sites by creating an optimized generation of our TRC 1-2 nuclease. While the first version of the nuclease produced modified CAR T cells that performed to specifications and were phenotypically normal, the optimized nuclease allowed for a large increase in the amount of cells produced during the manufacturing process and increased the effectiveness of the CAR T cells in killing the target cells. In this case, it appears that off-target genome breaks that occur during the production of cell therapy products may have a greater effect on the efficiency and cost of manufacturing than the safety of the final product.

REFERENCES

1. Tsai, S. Q., Zheng, Z., Nguyen, N. T., Liebers, M., Topkar, V. V., Thapar, V., ... Joung, J. K. (2015). GUIDE-Seq enables genome-wide profiling of off-target cleavage by CRISPR-Cas nucleases. *Nature Biotechnology*, 33(2), 187-197.

2. MacLeod, D. T., Antony, J., Martin, A. J., Moser, R. J., Hekele, A., Wetzel, K. J., ... & Bartsevich, V. V. (2017). Integration of a CD19 CAR into the TCR alpha chain locus streamlines production of allogeneic gene-edited CAR T cells. *Molecular Therapy*, 25(4), 949-961.